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SYNTHESIS AND PHARMACOLOGICAL PROFILE OF NEW 1, 3-DISUBSTITUTED CYCLOHEXANES AS LEUKOTRIENE B4 RECEPTOR ANTAGONISTS

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Abstract: In the course of developing stable leukotriene B₄ antagonists, we synthesized a novel non aromatic series of compounds containing a 1, 3-disubstituted cyclohexane ring in place of the conjugated double bonds of the natural eicosanoid. The Structure-Activity Relationship (SAR) studies leading to the identification of the acid 1d are described. Copyright © 1996 Elsevier Science Ltd

Leukotriene B₄ (LTB₄) is a 5-lipoxygenase product of arachidonic acid metabolism which stimulates human polymorphonuclear leukocyte (PMN) functions. LTB₄ is a potent inducer of neutrophil chemotaxis¹, aggregation², degranulation³ and respiratory burst⁴ in vitro. It has been postulated to be a major inflammatory mediator⁵ in a number of disease states such as rheumatoid or spondyloarthritis⁶, psoriasis⁷, ulcerative colitis⁸ and some respiratory diseases⁹. Consequently, a number of potent and selective LTB₄ receptor antagonists have been developed in the past few years, the large majority of them being aromatic compounds¹⁰ Both LY-223982¹¹ and SC-41930¹² are structurally related to the LTD₄ receptor antagonist FPL-55712, while SM-9064¹³ and U-75302¹⁴ are related to LTB₄, but exhibit partial agonist activity. Recent reports described the high affinity of ONO 4057¹⁵ and RG-14893¹⁶, which are pure antagonists.

As part of our studies directed towards the identification of LTB₄ antagonists based on the structure of the natural ligand itself, we thought it was of interest to check the hypothesis that designed cyclohexylic compounds could mimic the conjugated double bonds of leukotriene B₄. This concept offered numerous opportunities for obtaining analogs with restricted conformational freedom, with the goal of identifying a stable LTB₄ receptor antagonist.

During the course of that research, compound $\underline{1a}$ was identified as an early development 17 and became the focus of SAR studies. We wish to describe here our initial efforts in this area with the synthesis of the diastereoisomer compounds $\underline{1}$, $\underline{2}$, $\underline{3}$, and $\underline{4}$ and the isolation of the enantiomers of $\underline{1d}$ ($\underline{1d1}$ and $\underline{1d2}$).

Chemistry:

The general synthetic route to compounds $\underline{1-4}$ is illustrated in schemes I and II¹⁸. The various ω chains of these compounds were introduced in good yields on the cyclohexylic ring by Horner-Wadsworth-Emmons reactions ¹⁹ between the crude unstable aldehyde $\underline{2}$ and the ylides of the suitable β -ketophosphonates (Scheme I). The required methyl 3-oxo-cyclohexanecarboxylate $\underline{7}$ was prepared ²⁰ by successive hydrogenation and oxidation of methyl 3-hydroxy benzoate $\underline{5}$. The reduction of the protected keto-ester $\underline{8}$ with diisobutylaluminium hydride at -78°C was monitored by Gas Chromatography in order to control the formation of alcohol (over reduction). As soon as traces of alcohol appeared (after 30 min), the resulting crude aldehyde $\underline{9}$ was added on the ylide of the β -ketophosphonate (NaH, THF) at -40°C. ¹H NMR proved the enones $\underline{10}$, obtained in 75-85% yields, to be only of E configuration ²¹, no trace of Z analogue being detected.

Scheme I

OH
$$CO_{2}CH_{3}$$

$$C$$

Keys: (a) $R_1 = (CH_2)_4CH_3$ (b) $R_1 = (CH_2)_4CH_3$ (c) $R_1 \approx (CH_2)_7CH_3$ (d) $R_1 = (CH_2)_4Ph$. Reagents and conditions (isolated yield):

(i) H_2/Ru , 100 bars, 120°C, ethanol (93%); (ii) CrO_3 , H_2SO_4 (82%); (iii) Ethylene glycol, APTS, toluene reflux (95%); (iv) DIBAL-H, toluene, -78°C; (v) β -ketophosphonate, HNa, -40°C (70-86%); (vi) NaBH₄, CeCl₃, methanol (92-99%); (vii) SiO₂, H_2SO_4 (76-77%); (viii) TBDMSCl, DBU, CH_2Cl_2 (85-89%).

These enones were then reduced to the corresponding enols 11 by treatment with sodium borohydride in the presence of cerium chloride²², in order to avoid simultaneous hydroboration of the double bond. After deprotection of the carbonyl function²³, the allylic alcohols 12 were protected as their *tert*-butyldimethylsilyl ethers 13. This key intermediate synthon allowed the homologation of a large variety of α chains by 1,2-additions on the carbonyl function (Scheme II). The condensation of the ethyl acetate carbanion (LDA at -78°C) afforded compounds 1-4 b, c, d, while the four carbons-homologated compounds 1-4 a were obtained using the previously described lithiated OBO ortho-ester²⁴. In each series (a to d) the 14 trans and 14 cis silylated ethers were separated by column chromatography, and their relative trans and cis configurations established by ¹H NMR²⁵. As expected, the bulky group was locked in equatorial position. Deprotection of 14 trans and 14 cis hydroxy-esters, followed by careful preparative HPLC separation provided respectively and by order of elution the pure 2-1 and 4-3 diastereoisomers²⁶. At this stage of the research, all the racemic compounds 1-4 a-d were converted into their sodium salts (NaOH, methanol) for biochemical/pharmacological testing. The racemic mixture of compound 1d was resolved by chiral HPLC²⁷ to afford successively by order of elution the enantiomers 1d1 and 1d2.

Scheme II

HO,
$$R_1$$
 (ix)

HO, R_2
 R_1
 R_1
 R_2
 R_2
 R_1
 R_2
 R_2
 R_1
 R_2
 R_2
 R_2
 R_1
 R

(c) $R_1 = (CH_2)_7 CH_3$ $R_2 = CH_2 CO_2 Et$; Reagents and conditions (isolated yield):

(a) $R_1 = (CH_2)_4CH_3$ $R_2 = (CH_2)_3CO_2H$;

Keys:

(ix a) OBO ortho ester, tBuLi, THF, -78°C (56%) or (ix b,c,d) Ethyl acetate, LDA, THF, -80°C (87-92%); (x) HCl 1N, THF (82-84%).

(b) $R_1 = (CH_2)_4 CH_3$

(d) $R_1 = (CH_2)_4 Ph$

 $R_2 = CH_2CO_2Et$;

 $R_2 = CH_2CO_2Et$

Biological results:

The diastereoisomers 1 a-d to 4 a-d were tested for their ability to inhibit the binding of LTB₄ to human neutrophil membranes²⁸. In each series (a to d), the more polar 1 of the *trans* diastereoisomers 2 and 1 exhibited the highest competitive effect (Table I), compared to compounds 2, 3 and 4, which also competed for [³H] LTB₄ binding but with lower affinities (not shown).

Table I: Inhibition of specific binding of [3H] LTB₄28 to human neutrophils

$$R_2$$
 R_1
 R_2
 R_1

| Compound | R_1 | R ₂ | IC ₅₀ (μM) ^a | RBA b |
|------------|---|--|------------------------------------|--------|
| <u>1a</u> | (CH ₂) ₄ CH ₃ | (CH ₂) ₃ CO ₂ Na | 70 | |
| <u>1b</u> | (CH2)4CH3 | CH ₂ CO ₂ Na | 8 | 1/1000 |
| <u>1c</u> | $(CH_2)_7CH_3$ | CH ₂ CO ₂ Na | 1 | 1/125 |
| <u>1d</u> | (CH ₂) ₄ Ph | CH ₂ CO ₂ Na | 0.8 | 1/100 |
| <u>1d1</u> | $(CH_2)_4Ph$ | CH ₂ CO ₂ Na | 0.3 | 1/40 |
| <u>1d2</u> | $(CH_2)_4Ph$ | CH ₂ CO ₂ Na | 70 | |

a IC₅₀ are extrapolated from mean competition curves obtained from at least three different experiments.

Shortening of the carboxylic chain (<u>1a</u> to <u>1b</u>), lenghtening of the lipidic tail (<u>1b</u> to <u>1c</u>) and introduction of a phenyl group at the end of the lipophilic chain, in order to obviate ω -oxydation²⁹ as for LTB₄³⁰, led to the stable compound <u>1d</u> which exhibited substantial affinity for LTB₄ receptor (RBA=1/100). The first eluted enantiomer of <u>1d</u> in chiral HPLC (<u>1d1</u>) elicited as expected a higher affinity than the racemic <u>1d</u>, the other enantiomer (<u>1d2</u>) only demonstrating a very weak affinity for the receptor.

The antagonist properties were then evaluated against LTB₄-induced human neutrophils chemotaxis³¹ and guinea pig lung parenchyma contraction³². It was very encouraging that none of the four isomers 1-4 demonstrated any significant LTB₄ receptor agonist activity at concentrations up to 50 μM, in opposition with U-75302¹⁴, SC-45694³³, and potent disubstituted pyridines analogues³⁴, which were found to be partial functional agonists for LTB₄ receptor.

Compound <u>1d</u> inhibited the LTB₄-induced chemotaxis (pKB: 6.5) and the contractile effect of LTB₄ on guinea pig lung parenchyma strips (IC₅₀: 40nM). In the PMA (phorbol 12-myristate 13-acetate)-induced ear oedema test³⁵, <u>1d</u> exhibited a potent anti-inflammatory activity when topically applied on the skin (EC₅₀ = 10

b RBA: Relative binding affinity: IC₅₀ LTB₄ / IC₅₀ compound.

μg/ear). This effect is likely related to LTB₄ antagonism, since <u>1d</u> has been shown not to modify either cyclooxygenase or lipoxygenase activities³⁶ at the highest tested concentration (10⁻⁵ M).

In conclusion the replacement of the unstable triene unit of the natural eicosanoid for a cyclohexylic ring afforded compounds eliciting good affinities for the LTB₄ receptor and interesting antagonist activities. The designed compounds being stable rigid mimics of the natural ligand, they may provide useful tools which could lead to significant advances in our knowledge of the tridimensional structure of the receptor itself.

Finally, in this new series of 1, 3-disubstituted cyclohexane compounds, the resolution of the racemic mixture of compound <u>1d</u> afforded one enantiomer (<u>1d1</u>) being 200 fold more active than the other one (<u>1d2</u>). Work is currently under investigation in order to determine the absolute configuration of the enantiomerically pure <u>1d1</u> and try to correlate it to the known configuration of LTB₄. This attractive compound has been selected for further development and is currently subjected to functional modifications.

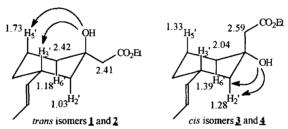
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References and Notes:

- 1. Ford-Hutchinson, A.W.; Bray, M.A.; Doig, M.V.; Shipley, M.E., Smith, M.J.H. Nature 1980, 286, 264.
- 2. Ringertz, B.; Palmblad, J.; Radmark, O.; Malmsten, C. FEBS Lett. 1982, 147, 180.
- 3. Rae, S.A., Smith, M.J.H. J. Pharm. Pharmacol. 1981, 33, 616.
- 4. Flohé, L.; Gunzler, W.A.; Loschen, G.; Günzler, W. A. In Superoxide and superoxide dismutase in chemistry, biology and medecine; Sies, E., Ed.; Elsevier Science Publishers: Amsterdam, 1986, p 376
- 5. Camp, R.D.R.; Coutts, A.A.; Greaves, M.W.; Kay, A.B.; Walport, M. J. Br. J. Pharmacol. 1983, 80, 497
- Klickstein, L.B.; Spagleigh, C.; Goetzl, E. J. J. Clin. Invest. 1980, 66, 1166.
- 7. Brain, S.; Camp, R.; Dowd, P.; Kobza-Black, A.; Greaves, M. J. Invest. Dermatol. 1984, 83, 70
- 8. Sharon, P.; Stenson, W. F. Gastroenterology 1984, 86, 453.
- 9. O'Driscoll, B.R.; Cromwell, O.; Kay, A.B. Clin. Exp. Immunol. 1984, 55, 397.
- 10. Daines, R.A.; Chambers, P.A.; Eggleston, D.S.; Foley, J.J.; Griswold Don, E.; Haltiwanger, R.C.; Jakas, D.R.; Kingsbury, W.D.; Martin, L. D.; Pendrak, I.; Schmidt, D.B.; Tzimas, M.N.; Sarau, H. J. Med. Chem. 1994, 37, 3327.
- 11. Gapinski, D.M.; Mallett, B.E.; Froekich, L.L.; Jackson, W.T. J. Med. Chem. 1990, 33, 2798.
- 12. Djuric, S.W.; Collins, P.W.; Jones, P.H.; Shone, R.L.; Tsai, B.S.; Fretland, D.J.; Butchko, G.M.; Villani-Price, D.; Keith, R.H.; Zemaitis, J.M.; Metcalf, L.; Bauer, R.F. J. Med. Chem. 1989, 32, 1145.
- 13. Namiki, M.; Iganashi, Y.; Sakamato, K.; Koga, Y. Biochem. Biophys. Res. Commun. 1986, 138, 540.
- 14. Morris, J.; Wishka, D.G. Tetrahedron Lett. 1988, 29, 143.
- (a) Konno, M.; Sakuyama, S.; Nakae, T.; Hamanaka, N.; Miyamoto, T.; Kawasaki, A. Adv. Prostaglandin, Thromboxane & Leukotriene Res. 1991, 21, 411. (b) Kashikawa, K.; Tateishi, T.; Maruyama, T.; Seo, R.; Toda, M.; Miyamoto, T. Prostaglandins 1992, 44, 261.
- Huang, F.; Chan, W.; Warus, J.D.; Morrissette, M.M.; Moriarty, K. J.; Chang, M.N.; Travis, J.J.; Mitchell, L.S.; Nuss, G. W.; Sutherland, C.A. J. Med. Chem. 1992, 35, 4253.
- 17. Hullot, P.; Poudrel, J.M.; Escale, R.; Girard, J.P.; Rossi, J.C.; Muller, A.; Bonne, C.; 9th International Conference on Prostaglandins and related Compounds, Florence, 1994, June 6-10, Abstract p. 43.

- 18. All new compounds exhibited satisfactory ¹H and ¹³C NMR, IR, elemental analysis and/or high resolution mass spectra data. Diastereomeric mixtures of compounds <u>1</u>, <u>2</u>, <u>3</u> and <u>4</u> were not separated, except <u>1d</u>.
- 19. Takacs, J.M.; Helle, M.A.; Seely, F.L. Tetrahedron Lett. 1986, 27, 1257.
- 20. House, H.O.; Latham, R.A.; Slater, C.D. J. Org. Chem. 1966, 31, 2667.
- 21. Compound 10: ¹H NMR (CDCl₃, 360 MHz), J_{trans} = 15.9 Hz.
- 22. Gemal, A.L; Luche, J.L. J. Am. Chem. Soc. 1981, 103, 5454.
- 23. Huet, F.; Lechevallier, A.; Pellet, M.; Conia, J.M. Synthesis 1978, 63.
- Kugel, C.; Lellouche, J.P.; Beaucourt, J.P.; Niel, G.; Girard, J.P.; Rossi, J.C. Tetrahedron Lett. 1989, 30, 4947.
- 25. The respective *cis* and *trans* configurations were easily attributed, using the influence of the hydroxyl group position on the axial protons chemical shifts in ¹H NMR (CDCl₃, 360 Mhz).



Due to the field effect of the oxygen atom, the axial cyclohexylic protons H₅ and H₃ are more deshelded when the hydroxyl function is loocked in axial position.

- 26. Spectral data for compound <u>1d</u>: I.R.: 3410, 1720, 1185, 960; ¹H-NMR (CDCl₃): 0.85 1.80 (m; 14H); 1.26 (t; 3H; J = 7.1 Hz); 2.41 (s; 2H); 2.42 (m; 1H); 2.59 (t; 2H; J = 7.7 Hz); 4.00 (q; 1H; J = 6.4 Hz); 4.16 (q; 2H; J = 7.1 Hz); 5.40 (dd; 1H; J = 15.5 Hz, J = 6.7 Hz); 5.52 (dd; 1H; J = 15.5 Hz, J = 6.4 Hz); 7.15 (m; 3H); 7.25 (m; 2H);
- 27. Column used: Chiralcel[®] OD (Daicel), 20 μm, 250×50 mm. UV detection at 210 nm. Elution (100 ml/min) with heptane/isopropanol (90/10) afforded successively the enantiomers <u>1d1</u> (13 min) and <u>1d2</u> (21 min).
- 28. Lin, A.H.; Ruppel, P.L.; Gorman, R.R. Prostaglandins 1984, 28, 837.
- 29. Miller, W.L.; Weeks, J.R.; Lauderdale, J.W.; Kirton, K.T. Prostaglandins 1975, 9, 9.
- 30. Hansson, G.; Lindgren, J.A.; Dahlens, S.E.; Hedquist, P.; Sammuelsson, B. FEBS Lett. 1981, 130, 107.
- 31. Isolated human neutrophils labelled with ⁵¹Cr (1 µCi/10⁶ cells) for 1h at 37°C, were suspended at 10⁷ cells/ml in Hank's buffer supplemented with 1% bovine serum albumin. LTB₄ and competitors in Hank's buffer were added to the bottom half of Boyden-Keller chambers; two 3 µm cellulose nitrate filters were placed over each well and the top part of the chambers was filled with cell suspension. The chambers were incubated at 37°C for 150 min. Radioactivity on the lower filter was measured by liquid scintillation spectrometry.
- 32. Strips of Dunkin Hartley guinea pig lung parenchyma were placed in organ baths containing oxygenated Tyrode solution at 37°C. Strips were recorded on a polygraph with an isometric tension of 400 mg. After a 1h equilibration mepyramine (1μM) and atropine (10 μM) were added to the Tyrode solution and the compound was tested for its agonist activity. The antagonist activities were tested by adding the drugs to the bath 2 min before LTB₄ (30 nM) stimulation.
- Tsai, B.S.; Keith, R.H.; Villani-Price, D.; Haack, R.A.; Djuric, S.W. J. Pharm. Exp. Ther., 1994, 268, 1493.
- 34. Kingsbury, W.D.; Pendrak, I.; Leber, J.D.; Boehm, J.C.; Mallet, B.; Sarau, H.M.; Foley, J.J.; Schmidt, D.B.; Daines, R.A. J. Med. Chem. 1993, 36, 3308.
- 35. Inoue, H.; Mori, T.; Shibata, S.; Koshihara, Y. Br. J. Pharmacol. 1989, 96, 204.
- 36. Evaluations of cycloxygenase/lipoxygenase activities have been performed by Fournier and Servier Pharmaceutical Companies, according to the methods described by Bonne, C.; Muller, A.; Latour, E.; Tissie, G; Emerit, I.; Modat, G.; Dornand, J.; Roche, M.; Giroud, J.P.; Griswold, D.E.; Marshall, P.J., Coquelet, C. Arzneim. Forsch., Drug Res. 1989, 39, 1242.